

World Headquarters

Millar, Inc.
6001-A Gulf Freeway
Houston, Texas 77023 USA
Phone: 832.667.7000 or 800-669-2343 (in the USA)
Fax: 713-714-8497
Email: info@millar.com
Web site: millar.com

Millar Worldwide Distribution

Millar, Inc. has a network of Authorized Distributors in most countries around the world. For information on the Millar distributor in your country, please contact the Millar Customer Service Department at our headquarters in Houston, Texas.



Making the
improbable possible.

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M.I. P/N: 004-2102 Rev. H

**Mikro-Tip® Ultra-Miniature
Pressure-Volume Catheter****Animal Use Only****Instructions for Use**

Service Provision

Consult web site below for service information:

www.millar.com

Millar Limited Warranty

Millar warrants that at the time of sale to the original purchaser, the device shall be free from defects in materials and workmanship for a period of six (6) months from its date of shipment to the original purchaser. If there is such a defect, Millar will, at no charge and at its option, either repair or replace any Mikro-Tip P-V catheter as appropriate. Millar's limited warranty does not cover damage to the product from alterations, misuse, abuse, negligence, or accident.

The user shall determine the suitability for use of these devices for research purposes only. Therefore, the user accepts these devices subject to all the terms hereof. Furthermore, Millar does not warrant that the equipment is suitable for any specific purpose, other than that explicitly stated by Millar.

Furthermore, Millar does not warrant its Mikro-Tip P-V catheter for use with signal conditioning hardware that is not manufactured by Millar. Using the Mikro-Tip P-V catheter with signal conditioning hardware that is not manufactured by Millar will render the warranty null and void and the catheters will not be considered for evaluation or repair.

In addition, Millar makes no warranty regarding device efficacy after three (3) years from the date of manufacture. Catheters beyond the age of three (3) years will not be considered for evaluation or repair.

This warranty is in lieu of and excludes all other warranties not expressly set forth herein, whether expressed or implied warranties of merchantability or fitness of purpose. Since handling, storage, cleaning and sterilization of the product, as well as factors relating to catheterization procedures, and other matters beyond Millar's control directly affect the product and the results obtained from its use, Millar shall not be liable for incidental or consequential loss, damage, or expense directly or indirectly arising from the use of this product.

Figures

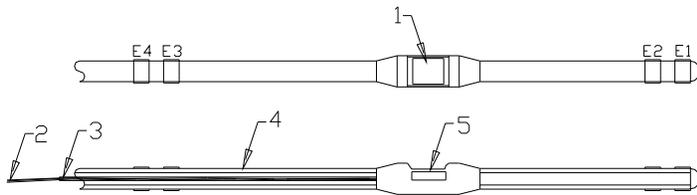


Figure 2

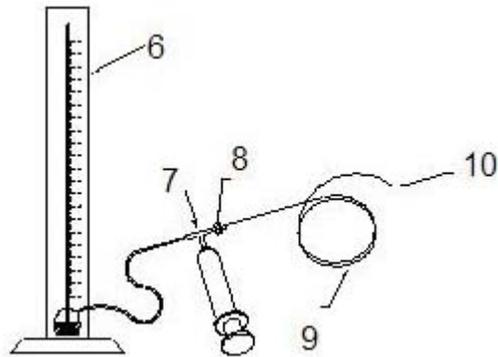


Figure 3

Figure Legend

- | | |
|------------------------------|---------------------------------|
| 1. Pressure Sensing Area | 6. Mercury Manometer |
| 2. Wires to Connector | 7. Tee Fitting |
| 3. Vent to Connector | 8. Plastic Dome |
| 4. Catheter | 9. Pressure Transducer Catheter |
| 5. Silicone Rubber Diaphragm | 10. To Recorder |

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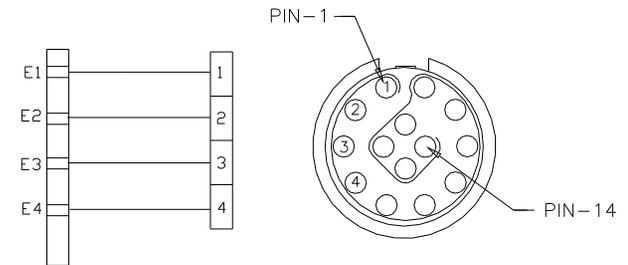
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Recommended Accessories

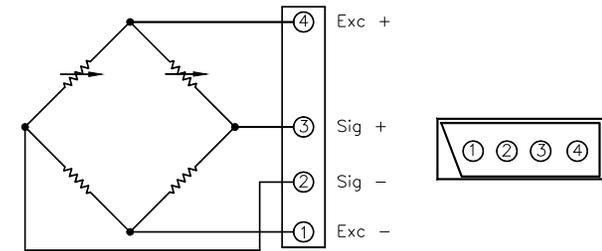
M.I. P/N 880-0168, MPVS Ultra
 M.I. P/N 880-0168SS, MPVS Ultra Single Segment
 M.I. P/N 850-5103, PEC-4D Pressure Cable
 M.I. P/N 850-5099, CEC-4B Volume Cable
 M.I. P/N 880-0160, MPVS-300
 M.I. P/N 880-0159, MPVS-400
 M.I. P/N 850-5103, PEC-4D Pressure Cable
 M.I. P/N 850-5099, CEC-4B Volume Cable
 M.I. P/N 910-1048, Large Volume Calibration Cuvette
 M.I. P/N 910-1049, Small Volume Calibration Cuvette
All accessories sold separately.

Definition of Symbols	
	Attention, consult accompanying documents
	Date of Manufacture
REF	Catalog Number
SN	Serial Number
	Batch Code
	Use By Date
	Electrostatic Sensitive Device
	EU Declaration of Conformity

Schematics



Electrode & Redel Connector



Sensor & Low Profile Connector

Device Description

The Mikro-Tip Pressure-Volume (P-V) catheter combines an ultra-miniature pressure sensor(s) and a series of electrodes mounted at various locations along the distal end of the catheter (Figure 2). The catheter terminates at two electrical connectors at the proximal end. The pressure sensor produces an electrical output signal, which varies in direct proportion to the magnitude of sensed pressure. The electrodes allow for estimation of left ventricular volume changes by impedance measurements.

Mikro-Tip catheters are intended for multiple uses. Experience has proven that the instruments are safe and effective for extended service if proper handling, cleaning, and sterilization procedures are followed.

Immediately upon receipt of the catheter, and prior to its initial cleaning, sterilization, and use, the customer should verify that the catheter is operational.

Sensor Specifications

		1.4 - 2F Catheter	1F Catheter
Type of Sensor		Diffused Semiconductor, piezoresistive	
Pressure Range		-50 to + 300 mmHg (-6.7 to 40 kPa)	
Overpressure		+4000 mmHg (+530 kPa), -760 mmHg (-100 kPa)	
Rated Excitation*		2.5-7.5 V _{DC} or V _{AC} rms	2.0-10.0 V _{DC} or V _{AC} rms
Sensitivity		5 μV/V/mmHg, nominal (37.6 μV/V/kPa)	
Temperature Error Band at Zero Pressure		±3 mmHg (± 0.4 kPa) BSL, 23 - 38 °C	
Linearity and Hysteresis (Combined)		±1%, BSL of full scale	±2%, BSL of full scale
Drift**		< 6 mmHg (0.8 kPa) in 12 hours	
Natural Frequency		≥ 10 kHz	
Bridge Resistance		1000 ohms, nominal	
Reference Pressure		Atmosphere	
Electrical Leakage	Single Pressure	< 10 μA at 180 V _{DC} < 10 μA at 120 V _{AC}	< 10 μA at 25 V _{DC}
	Double Pressure	< 10 μA at 25 V _{DC}	< 10 μA at 25 V _{DC}
Zero Offset		< ±50 mmHg (± 6.7 kPa)	

* Performance specifications are for 5 V_{DC}. Transient voltages up to 20 volts will not damage the transducer.

** Based on 30 minute presoak.

Catheter Specifications

Pin-to-Electrode Resistance: <15 ohms

Catheter Size: Per package labeling

Usable Length: Per package labeling

Electrodes: 0.25 mm

Intended Use/Indications

Use of the ultra-miniature Mikro-Tip P-V catheter is indicated when combined physiological pressure and volume (as estimated by electrical impedance measurements) monitoring is required in small animals such as mice and rats.

Mikro-Tip P-V catheters may be introduced into the left ventricle using a closed chest carotid artery approach or using an open chest approach through an entry site made in the apex of the heart with a needle.

Proper catheter positioning within the left ventricle can be verified with the aid of echocardiography.

This product is designed for use by professionals with appropriate education and training in life science and medical research applications.

Warnings

- The Mikro-Tip P-V catheter should be stored in a cool, dry place.
- The Mikro-Tip P-V catheter is shipped with a foam dome fitting over the catheter tip to protect the sensor/electrode area. The catheter tip should be left in the foam dome when the catheter is in the tray.

Precautions

Proper handling of the Mikro-Tip P-V catheter is critical to avoid damage and maximize the catheter's use-life.

- Use of the Mikro-Tip P-V catheter should be restricted to specialists who are familiar with, and have been trained to perform, the catheterization procedures for which the device is intended.
- Do not under any circumstances bend, kink, fold, cut, pinch, crush, etc. the Mikro-Tip P-V catheter especially around the sensor/electrode area. These actions could severely damage the catheter and/or render the catheter unusable and result in costly repair or replacement charges.
- The Mikro-Tip P-V catheter should be inspected for damage (cracking, kinks, etc.) prior to each use.
- Do not grip the Mikro-Tip P-V catheter with forceps, tweezers, or fingers anywhere near the sensor/electrode area.
- When using metal forceps or tweezers to grip the Mikro-Tip P-V catheter, slip some soft tubing over the tips of the metal instrument to cushion the interface between the instrument and the catheter body.
- Always grip the Mikro-Tip P-V catheter proximal to the sensor/electrode area. Never grip the catheter between the electrodes or between the electrodes and the pressure sensor case.
- Always be mindful of the location of the catheter tip.
- Do not set heavy objects or metal instruments on top of the Mikro-Tip P-V catheter.
- Do not tighten sutures over the sensor/electrode area on the Mikro-Tip P-V catheter.
- When using the carotid artery approach to insert the Mikro-Tip P-V catheter into the left ventricle, advance the sensor/electrode area completely beyond the proximal suture before tightening the suture.
- When using the carotid artery approach to insert the Mikro-Tip P-V catheter into the left ventricle, take care not to damage the catheter with the tips of any forceps or tweezers that are used to grip the artery when inserting the catheter.
- When using the open chest approach to insert the Mikro-Tip P-V catheter into the left ventricle, do not push the catheter tip directly through the left ventricular wall.

First, make an entry hole in the ventricular apex with a needle and then insert the catheter through the hole.

- Never apply excessive force when inserting the Mikro-Tip P-V catheter. The catheter may incur damage and could puncture through the left ventricular wall or damage the aortic valve.
- If resistance is encountered while inserting the Mikro-Tip P-V catheter, pull back slightly and then try advancing again.
- When removing the Mikro-Tip P-V catheter through the carotid artery, loosen the proximal suture before pulling the catheter out to insure the pressure sensor or electrodes do not catch on the suture and lift or tear away from the catheter.
- The Mikro-Tip P-V catheter should be cleaned immediately after each use (see Cleaning).
- The pressure sensor element on the Mikro-Tip P-V catheter is sensitive to electrostatic discharge. Refrain from touching the sensor element while the catheter is not connected to monitoring equipment.

Handling Precautions for Mikro-Tip P-V Catheters

	DO:	DO NOT:
Pressure Sensor & Electrodes	Clean immediately after use	Clean with stiff-bristled brush
	Protect with dome when not in use	Clean with high pressure water jet
	Disconnect during electrical defibrillation or electrosurgery	Tap the sensor or electrodes against a rigid surface
Catheter	Clean immediately after use	Apply excessive force to the sensor or electrodes
		Expose to excessive pressure
Connectors & Cables	Protect connectors from fluid	Cut, crease, knot, fold, kink, or crush with forceps or clamps of any kind
Cleaning	Keep catheter and sensor wet until cleaning	Immerse connectors in liquid
	Clean thoroughly with approved enzymatic cleanser immediately after use	Expose to alcohol, cresols, phenols, mercury compounds, hypochlorites, acetone, peroxide, silicone chlorine, xylenes, trichloroethylene, or freon
Disinfection or Sterilization	Dry catheter before sterilizing	Use ultrasonic cleaner.
	Remove plastic dome from catheter	Immerse electrical connector
		Autoclave, irradiate (gamma/e-beam), plasma, peroxide or formaldehyde vapor solutions
		Use Sporox or Cidex PA solutions

Method of Sterilization for Catheters and Extension Cables (Optional)

CAUTION: DO NOT sterilize by autoclaving, radiation (gamma or e-beam), plasma, peroxide or formaldehyde vapor solutions.

Catheters must be completely cleaned and dried before sterilization. Aerate at room temperature or in a heated aeration cabinet (max. 145 °F, 63 °C). Catheters may be sterilized in the white plastic shipping tray. The foam dome and connector caps must be removed and placed alongside the catheter inside the pouch during sterilization. The caps should be saved and reused each time the catheter is cleaned.

The tray, with lid, should be placed in a breathable polyethylene pouch (e.g., 3M™ Steri-Lok™).

CAUTION: The catheter should be completely dry before sterilization.

Ethylene Oxide Sterilization Cycle Parameters

Preheat phase:	Starting Temperature 110 °F (43°C) min. Duration 30 minutes
Initial Vacuum:	6.0 inHgA (20.3 kPa) Rate: 3 minutes
Nitrogen Flush: 2 cycles	
Nitrogen Addition to:	28.0 ± 0.5 inHgA (94.8 ± 1.7 kPa)
Rate:	1.4 ± 0.5 inHgA/min. (4.7 ± 1.7 kPa/min.)
Evacuation:	6.0 ± 0.5 inHgA (20.3 ± 1.7 kPa)
Rate:	1.0 ± 0.5 inHgA/min. (3.4 ± 1.7 kPa)
Conditioning	
Humidification:	1.5 ± 0.5 inHgA (5.1 ± 1.7 kPa)
Steam Conditioning:	10 min.
Humidity Dwell:	30 ± 5 min. at 7.5 ± 0.5 inHgA (25.4 ± 1.7 kPa)
Relative Humidity:	15-70%
Ethylene Oxide Concentration:	500 ± 50 mg/L
Dwell Pressure:	16.5 ± 1.0 inHgA (55.8 ± 3.4 kPa)
Dwell Time:	2 hours
Temperature:	110-130 °F (43-54 °C)
Relative Humidity:	30-70% (35-44% nominal)
After Vacuum	
Vacuum:	6.0 ± 0.5 inHgA (20.3 ± 1.7 kPa)
Rate:	1.0 ± 0.5 inHgA/min. (3.4 ± 1.7 kPa)
Vacuum Hold:	10 min.
Gas Wash A:	4 cycles (minimum)
Release:	30.0 inHgA/min. (94.8 ± 1.7 kPa)
Rate:	1.4 ± 0.5 inHgA/min. (4.7 ± 1.7 kPa)
Vacuum	6.0 ± 0.5 inHgA (20.3 ± 1.7 kPa)
Rate:	1.0 ± 0.5 inHgA/min. (3.4 ± 1.7 kPa)
Release (Filtered Air):	28.0 ± 0.5 inHgA (94.8 ± 1.7 kPa)
Rate:	2.0 ± 0.5 inHgA/min. (6.6 ± 1.7 kPa)
Aeration (Hot Cell)	
Duration:	At least 8 hours
Temperature:	110 ± 10 °F (43 °C)

CAUTION: The Mikro-Tip transducer should not be used earlier than 5 days after sterilization.

6. After the cleaning procedure is complete, return the Mikro-Tip P-V catheter to its original packaging for storage:
 - Return the catheter to the tray.
 - Protect the sensor/electrode area on the catheter tip by carefully placing it within the foam dome that came with the packaging,
 - Slide the tray inside the box.
 - Store the catheter in a cool, dry place until the next use.

General Instructions for Catheter Disinfection

1. The catheter must be cleaned, rinsed and dried prior to disinfection. Soil, debris, proteins, and water can interfere with the effectiveness of the following procedure, posing a risk to the patient and the user. Note that some disinfectants have a limited usable life after activation or opening the container. Failure to heed such warnings can inhibit the effectiveness of the disinfection process.
2. Prepare the disinfectant according to the manufacturer's instructions.
3. Submerge the catheter into the disinfectant up to the strain relief. Do not submerge the strain relief or the connector as it will damage the transducer and void the warranty.
4. Soak the transducer in the disinfectant at the temperature and time intervals listed.

Rinsing after Disinfection

1. Rinse the device by submerging all exterior disinfected surfaces in sterile pyrogen-free water. The volume of the water should be at least two gallons (7.6 liters) and the soak time should be at least one minute.
2. At least three separate rinses are required. Do not reuse any of the water used for rinsing since it will be contaminated with the disinfectant.

Troubleshooting and Corrective Action

Problem	Probable Cause	Corrective Action
Excessive pressure signal drift	Deposit of foreign material on the diaphragm of the pressure sensor.	Follow Cleaning Instructions. If problem persists, contact Millar.
Pressure sensor will not balance (zero)	Moisture in the connector, damage to wires in the catheter, or fractured strain gauge within pressure sensor.	Follow Operating Instructions or substitute a transducer known to be operating properly into the recording system. If problem persists, contact Millar.
Noisy or erratic volume signal	Damage to electrodes or electrode-wire bond ("open electrode").	Follow Operating Instructions or substitute a transducer known to be operating properly into the recording system. If problem persists, contact Millar.

Maintaining Device Effectiveness

Catheter Preparation

Presoak the catheter tip in saline or distilled water maintained at body temperature for at least 30 minutes prior to use. Only soak the catheter tip, do not submerge the catheter's electrical connectors, as doing so will damage the sensitive electronics housed inside!

The presoak helps prepare the pressure sensor diaphragm for the "wet" biological environment. Presoaking the catheter tip in body temperature fluid also helps prevent pressure signal drift and negative pressure recordings. The presoak can be done by gently inserting the catheter tip into a beaker or dish or through the tip of a syringe.

Routine Inspection

CAUTION: If damage is found during inspection, DO NOT use the catheter. Contact Millar Instruments or an authorized Millar distributor for assistance.

The Mikro-Tip P-V catheter should be thoroughly inspected under magnification before and after each use to determine its condition. Carefully examine the catheter for cuts, kinks, or creases. The active surface of the sensor and electrodes should be examined for any film, blood, or biological debris that has not been removed by cleaning. Any film, blood, or biological debris may cause short-term baseline drift and should be removed. Such removal should consist of a thorough soaking in an enzymatic cleaning solution (Terg-A-Zyme[®]) followed by a persistent and gentle wiping action along the sensor or electrodes with a moist tissue, gauze, or cotton tipped swab. The connectors should undergo visual inspection for corrosion or bad contacts. Liquid entering the connector(s) can cause electrical hazard, erratic operation, and corrosion.

Pressure Calibration

Each transducer is calibrated for a standardized sensitivity of 5µV/V/mmHg (37.6 µV/V/kPa). Since the transducer electrical calibration may be altered by the effects of variations in monitors, calibration should be confirmed with a pressure manometer to help guarantee the most accurate results.

To properly calibrate and balance the Mikro-Tip P-V catheter pressure transducer, follow the instructions for the Millar control unit being used (MPVS Ultra or MPVS-300/400 for more details please see the "Operating Instructions" section of this manual). To confirm calibration accuracy, compare the pressure output produced using the pressure manometer with the electrical output signal produced by the Millar control unit for the Mikro-Tip P-V catheter pressure transducer (see Figure 3 for more detail).

Volume Cuvette Calibration

The cuvette calibration method uses an actual blood sample to make a more accurate assessment of the left ventricular volume. The cuvette calibration method involves using insulator-type cuvettes of known diameter filled with heparin-treated blood.

Figure 1 illustrates the procedure for measuring volume using the conductance method when a four-electrode catheter is submerged in a cylindrical non-conductive tube filled with a conductive solution. Conductance/Volume readings can be obtained by submerging the catheter electrodes into blood-filled tubes of known diameter.

The cuvette calibration is recommended for researchers who are interested in obtaining a more accurate assessment of left ventricular volume. Researchers can enhance the accuracy of their volume measurements by coupling the acquired calibration data with their own mathematical models and/or physical simulations to convert the volume measurements derived from the Millar hardware (MPVS Ultra or MPVS-300/400) into what they believe are the true volumes present within the left ventricle.

The cuvette calibration can dramatically affect the quality of the volume measurements. Many factors must be addressed to make the cuvette calibration as accurate as possible. These factors include: maintaining the cuvette at body temperature, minimizing blood clotting in the cuvette, properly cleaning the cuvette before performing the experiment, and centering the catheter within the cuvette.

The volume measured by the conductance method when a four-electrode catheter is submerged into a cylindrical non-conductive (e.g. Plexiglas) tube is given in Figure 1.

- L = distance between electrodes 2-3:
Catheter Model PVR-1045, L = 4.5 mm
Catheter Model SPR-839, L = 4.5 mm
Catheter Model SPR-869, L = 6.0 mm
Catheter Model's SPR-838 & SPR-847, L = 9 mm
- The actual volume between electrodes 2-3 = $\pi * r^2 * L$
- r = inner radius of cylinder

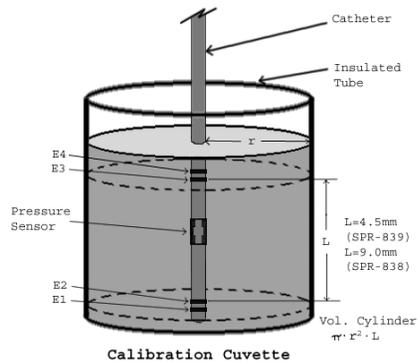


Figure 1.

The electronic μ S or RVU calibrator built into the Millar control unit (MPVS Ultra or MPVS-300/400) provides a quick and efficient way of calibrating the system. Units of conductance (RVU, μ mho, μ S) can be converted into volume units by following the cuvette calibration procedure listed below:

1. After the P-V loop data acquisition is complete, immediately remove the Mikro-Tip P-V catheter from the biological environment. When using the carotid artery approach, loosen the proximal suture before pulling the catheter back through the blood vessel. Loosening the proximal suture will help prevent the electrodes and pressure sensor from catching on the suture and breaking.
2. Upon removing the Mikro-Tip P-V catheter from the biological environment, immediately soak the catheter tip in a beaker, syringe, or dish filled with fresh saline or distilled water. Keep the tip soaking until you are ready to continue cleaning it or you are ready to catheterize another research subject. This step will help lyse any cells that adhere to the catheter and help loosen and prevent clot formations and protein build-up on the catheter tip.

NOTE

Never let blood dry on the catheter tip! Always keep the tip soaking in saline or distilled water between experiments or until you are ready to proceed with the next cleaning step.

Do not submerge the catheter connectors, as moisture will damage the sensitive electronics that are housed inside!

3. The Mikro-Tip P-V catheter should be thoroughly inspected under magnification after each use to determine its condition. Carefully examine the catheter for cuts, kinks, or creases. The active surface of the sensor and electrodes should be examined for any film or biological debris. Any film may cause short-term baseline drift and should be removed. Such removal should consist of a thorough soaking in Terg-A-Zyme (see below) followed by a persistent and gentle wiping action along the sensor or electrodes with a moist tissue, gauze or cotton tipped swab. The connectors should undergo visual inspection for corrosion or bad contacts.
4. Using an enzymatic cleaner on the distal portion of the Mikro-Tip P-V catheter is essential to preventing protein buildup on the pressure sensor and electrodes. Without the use of an enzymatic cleaner, a protein film will form on the pressure sensor that ultimately results in pressure signal drift. Millar provides the following guidelines for using enzymatic cleaners with the ultra miniature P-V catheters:
 - Use a Millar approved enzymatic cleaning agent such as Terg-A-Zyme by Alconox. www.alconox.com.
 - Follow the manufacturer's instructions for mixing the Terg-A-Zyme solution
 - Soak the catheter tip in a beaker or syringe for 15-30 minutes in the Terg-A-Zyme solution.
 - After the Mikro-Tip P-V catheter is done soaking in the enzymatic cleaning solution, thoroughly rinse the tip with fresh distilled water.
5. After the rinsing is complete, gently dry the Mikro-Tip P-V catheter according to the following steps:
 - Fold a Kim-wipe or similar soft tissue.
 - Use gentle stroking to brush dry the catheter tip.
 - Do not pull the catheter tip through a folded Kim-wipe.
 - Do not allow the catheter to air dry on the tray, table, or countertop.

samples/second) in order to accurately capture all of the features of the P-V waveforms produced by the fast beating hearts of mice and rats].

Cleaning

Approved Cleaners and Disinfectants

Type	Trade Name	Manufacturer	Active Ingredient.	Soak Time/Temperature
Enzymatic Detergent	Enzol® (in UK: Cidezyme®)	Advanced Sterilization Products (J&J)	Propylene Glycol	15 minutes / room temperature
	Endozime®	Ruhoff Corporation	Propylene Glycol	15 minutes / room temperature
	Terg-A-Zyme®	Alconox	Sodium Dodecylbenzene	15 minutes / room temperature
High-Level Disinfectant	Cidex Activated Dialdehyde Solution	Advanced Sterilization Products (J&J)	Glutaraldehyde	1-2 hours / 25 °C (77°F)
	Cidex® OPA	Advanced Sterilization Products (J&J)	Ortho-phthalaldehyde	16-30 minutes / 20 °C (68°F)
	MetriCide®	Metrex	Glutaraldehyde	1-2 hours / 25 °C (77°F)

DO NOT USE:

- Glutaraldehyde solutions containing surfactants (e.g., Cidex 7 or Cidex Plus 28 Day)
- Solutions containing hydrogen peroxide (e.g. Sporox)
- Cidex PA solution

Water Resistant Connector Caps

Each catheter has water-resistant caps to protect electrical pins and circuitry. Place caps over the open end of the connectors before cleaning and disinfecting. Remove caps prior to sterilization. Save and reuse these caps each time the catheter is cleaned and disinfected.

Cleaning Procedure

CAUTION: DO NOT submerge the wye junction or connectors. This will damage the catheter and void its warranty! Wipe with cleaner and gauze.

CAUTION: Use only the listed cleaners and disinfectants for the times/temperatures indicated.

CAUTION: Delays in rinsing greatly reduce cleaning effectiveness!

Procedure

1. In the data acquisition software, calibrate the hardware output using the electronic μS or RVU calibrator built into the Millar control unit (MPVS Ultra or MPVS-300/400). A linear two-point calibration is recommended at the available high and low μS points in the MPVS Ultra Control Software or 5RVU and 50RVU in the MPVS-300/400 (for more details on RVU calibration, please see the "Operating Instructions" section of this manual).
2. During the course of the experiment withdraw fresh slightly heparinized blood from the experimental subject and inject the blood into the different tubes of a calibration cuvette (insulated tubes with different diameters) to a depth that will submerge the Mikro-Tip P-V catheter electrodes when the catheter is dipped in the tube.

NOTE

Millar Instruments can provide standard calibration cuvettes for mice (Part No. 910-1049) and rats (Part No. 910-1048). Researchers who are interested in enhancing the accuracy of their volume calibrations may choose to fabricate their own cuvettes based on the size of their research animals.

3. Dip the Mikro-Tip P-V catheter in the center of the blood-filled tubes and record the conductance output in μS or RVUs. Keep the catheter centered in the tube (cuvette). If the bottom of the tube is not flat, then be careful to keep all four electrodes in the cylindrical portion of the tube.

NOTE

The blood used to perform the cuvette calibration must be free of saline or any other solution that may alter the conductance of the blood and affect the data unless you are developing calibration curves to help identify the changes in blood conductance that may be occurring as a result of introducing some type of compound, drug, etc.

4. Correlate the known volumes calculated from the catheter electrode spacing and the cuvette tubes with their respective conductance outputs to plot a graph of volume vs. conductance (μS or RVU).
5. Obtain the equation of the line (slope and y-intercept) and use it to convert conductance units (μS or RVU) to volume units (μL).

NOTE

As an alternative to a two-point calibration, some researchers follow the rules of good statistical sampling and choose to perform a multiple-point cuvette calibration of volume vs. conductance. This data can be recorded then plotted in a spreadsheet program such as Microsoft Excel to obtain the conductance (μS or RVU) to volume (μL) calibration. Please remember that the volume of blood within the animal as well as the size and number of calibration cuvettes used in the experiment limit the number of data points that can be acquired.

The cuvette calibration procedures listed above do not take into account the parallel volume contribution of the myocardium. Please consult Millar's MPVS Training CD hosted on Millar's Knowledge Center at millar.com/Knowledge-Center for more details on how to obtain a value for parallel volume.

Operating Instructions

1. Unpack the hardware, software, and cables.
2. Inspect the equipment for any obvious signs of external and/or internal damage. If anything is missing or the equipment seems to be damaged in any way, contact Millar Instruments or the authorized Millar distributor in your area immediately.
3. Turn on the computer that will be used to acquire and analyze the P-V data and install the necessary software (DAQ software with PV Loop analysis, Millar PVAN)
4. Connect the P-V channel outputs on the back panel of the Millar control unit (MPVS Ultra or MPVS-300/400) directly to a computer (USB cable for MPVS-400) or to the input channels on the data acquisition system using BNC-BNC cables or BNC cables that terminate at a connector that is compatible with the data acquisition system (MPVS Ultra or MPVS-300/400). Turn on the data acquisition system.
5. Connect the power cord to the power input jack on the back of the Millar control unit (MPVS Ultra or MPVS-300/400). Please ensure the power cord conforms to the standards in the country of operation. The end user may be responsible for providing his own power cords if the cords supplied by Millar are not sufficient for the country of operation. Cords must connect to a grounded power outlet for safety.
6. Turn on the Millar control unit (MPVS Ultra or MPVS-300/400). Ensure that the Power Indicator located on the front panel lights up.
7. Connect the pressure cable (PEC-4D or similar) and volume cable (CEC-4B or similar) to the appropriate input connectors on the back of the Millar control unit (MPVS Ultra or MPVS-300/400).
8. Unpack a Mikro-Tip P-V catheter, being careful to handle it with a high degree of care.
9. Connect the pressure transducer connector on the Mikro-Tip P-V catheter to the pressure cable (PEC-4D or similar).
10. Connect the conductance/volume transducer connector on the Mikro-Tip P-V catheter to the volume cable (CEC-4B or similar).
11. Using a shallow dish, beaker, or syringe, soak the catheter tip in body temperature distilled water or saline for at least 30 minutes prior to insertion into the biological environment.
12. For the MPVS-300/400, verify that the appropriate segment length has been selected using the Segment Select switch located on the front panel. For standard four-electrode P-V catheters, select the "Short" position. For six-electrode P-V catheters, select either the "Short" or "Long" position depending on the desired electrode spacing (segment length).
13. Open the data acquisition software on the computer.
14. Configure the data acquisition software according to user preferences so that it can effectively acquire, display, and record the P-V data produced by the Millar control unit (MPVS Ultra or MPVS-300/400).

15. *Calibration Setup:* Follow the instructions in the data acquisition software user's manual as well as Millar's online Knowledge Center or MPVS user's guide to perform two-point calibration procedures for both the pressure and volume channels. For simplified electronic calibration verification, use the MPVS Ultra Control Software or transducer/calibration controls on the front panel of the MPVS-300/400 to provide fixed voltage signal references for pressure (0-100 mmHg) and volume (in μS or 5-50 RVU) that may be used to quickly perform the two-point calibration procedure. Remember that RVU verification is only for Relative Volume Units, not absolute volume. Conversion of Relative Volume Units into true volume units requires additional steps (please refer to Millar's Knowledge Center for additional instructions regarding volume calibration techniques). Volume calibration for a given catheter may be performed using small tubes of known diameters (a calibration cuvette, which can be supplied by Millar Instruments) filled with fresh, slightly heparinized blood drawn from an experimental subject. Validation of the volume estimates provided by the Millar hardware can be performed by other means, e.g. by thermodilution, flow probes, hypertonic saline dilution, echo, etc.

CAUTION: The "zero" output produced by selecting 0 mmHg in the MPVS Ultra Control Software is an electrical zero, not an atmospheric zero

16. *Pressure Transducer Balancing:* After the catheter tip has soaked for at least 30 minutes, select Transducer in the MPVS Ultra Control Software under the Catheter Calibration tab. For the MPVS-300/400 set the Transducer/Calibration switch on the front panel of the Millar control unit to Transducer. Use the transducer balance arrows in the Control Software or unlock and adjust the Balance control knob for the pressure channel the catheter is connected to. Use the controls to adjust the pressure transducer output while dipping the transducer just below the surface of distilled water or saline (essentially a zero pressure environment). Adjust the controls until the output signal reads approximately 0 mmHg (as displayed by an indicator in the data acquisition software or similar monitor). and select the Balance Locked button or lock the knob into position.

CAUTION: Do not submerge the electrical connectors on the catheter.

For best stability and zero adjustment, shield the sensor from bright ambient light during the balancing procedure and allow the catheter-tip to soak in body temperature sterile water or saline for 30 minutes prior to insertion into the biological environment. Zero balancing of the pressure transducer should be done prior to each catheterization procedure.

NOTE

A pressure manometer may also be used to check the zero balance of the transducer and calibrate the pressure output at two points. To pressurize the transducer, a manometer can be connected to one end of a hemostasis valve or Toughy-Borst/Luer adapter with flexible tubing. Carefully slip the catheter through the other end of the valve/adaptor and seal the valve/adaptor around the catheter shaft proximal to the sensor/electrode area. See Figure 3 for more detail.

17. The system is now ready to begin conducting P-V experiments. Prepare the experimental subject for catheter insertion according to the lab's established experimental protocols. When the experimental subject is ready, insert the Mikro-Tip P-V catheter into the ventricle and begin recording P-V data [**Please Note: Millar Instruments recommends a data acquisition sampling rate of at least 1 kHz (1000**